

Welcome

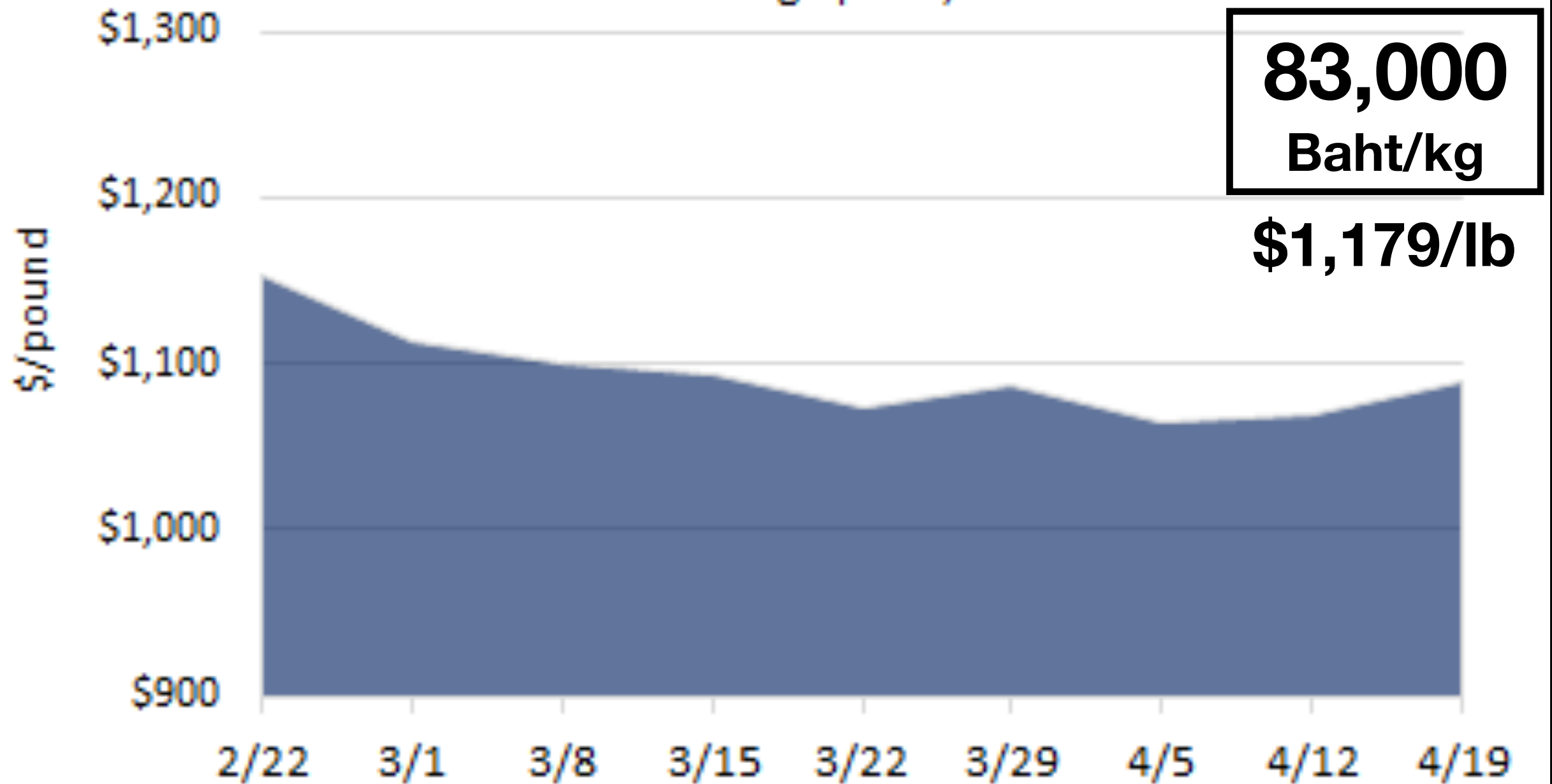


Cannabis & Cancer What's NEXT?

Somchai Thanasitthichai MD, FRCST

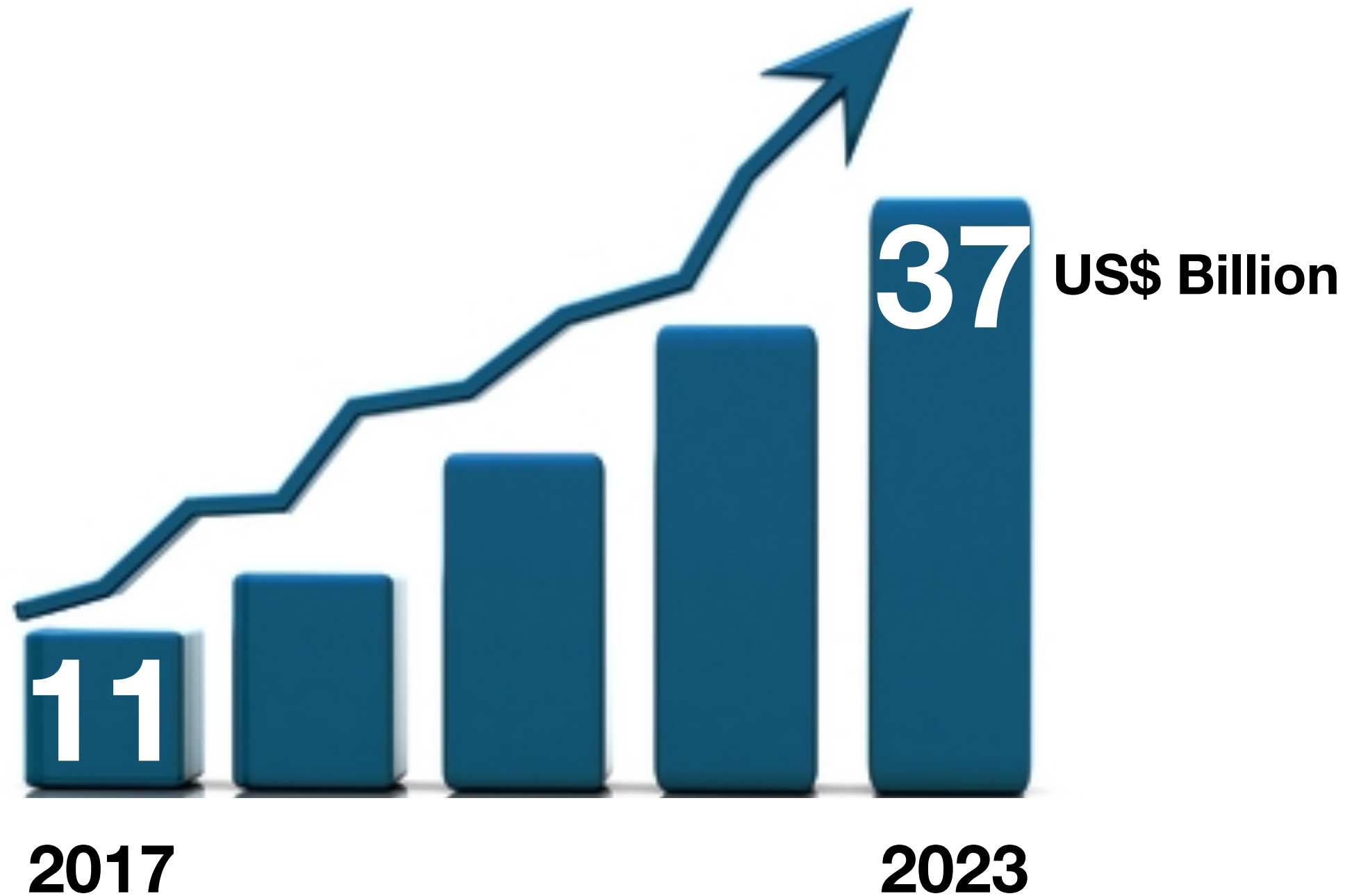
Deputy Director National Cancer Institute

Cannabis Benchmarks® US Spot Index
Week Ending Apr 19, 2019



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Global Medical Cannabis Market



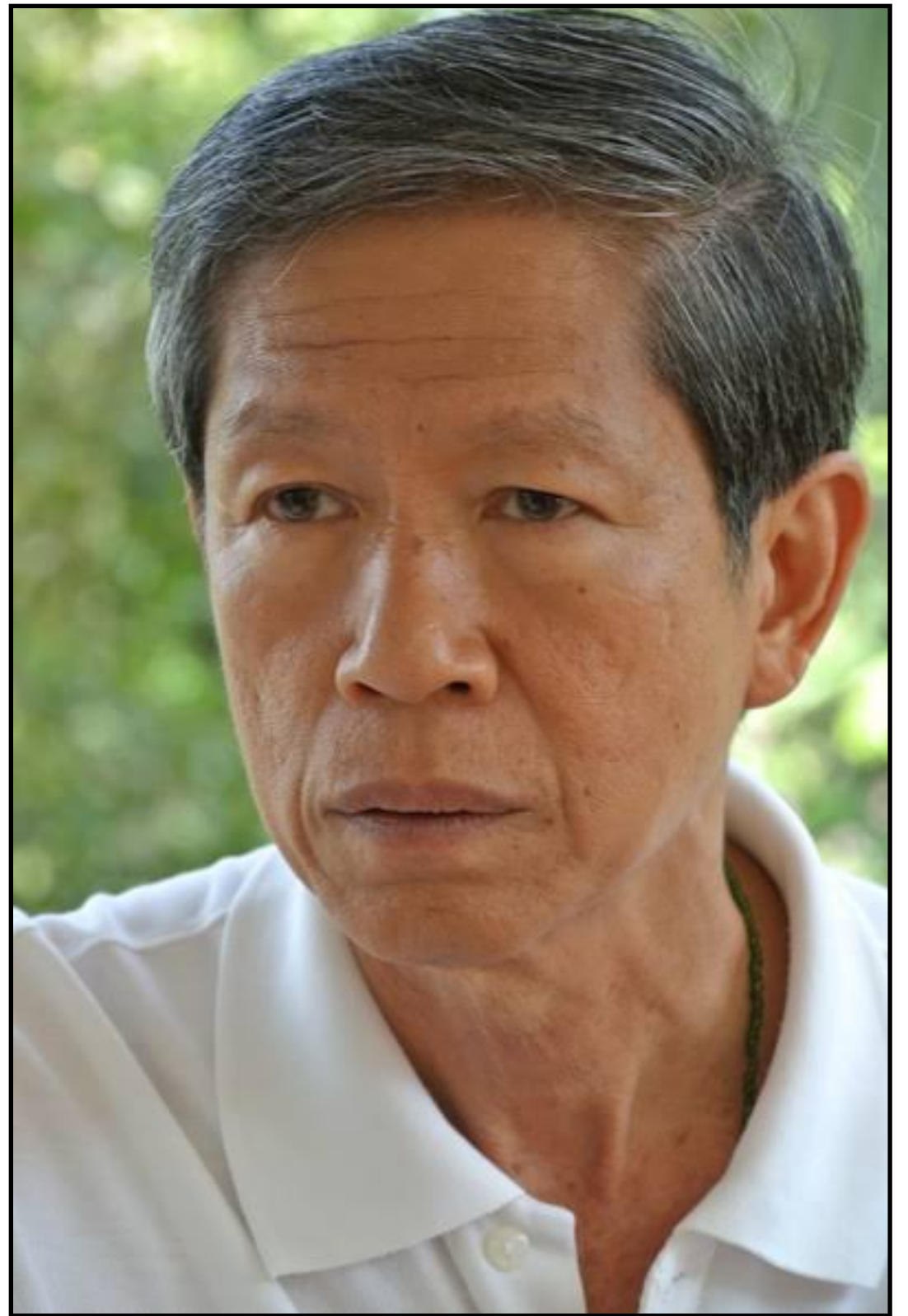
**‘When dealing with people,
remember you are not dealing
with creatures of logic, but
creatures of emotion’**

-Dale Carnegie



**“การแจกจ่ายกัญชา
เพื่อรักษาผู้เจ็บป่วย
เป็นเรื่องศีลธรรมที่ต้องทำ และ
อยู่เหนือกฎระเบียบบล้ำหลังใดๆ”**

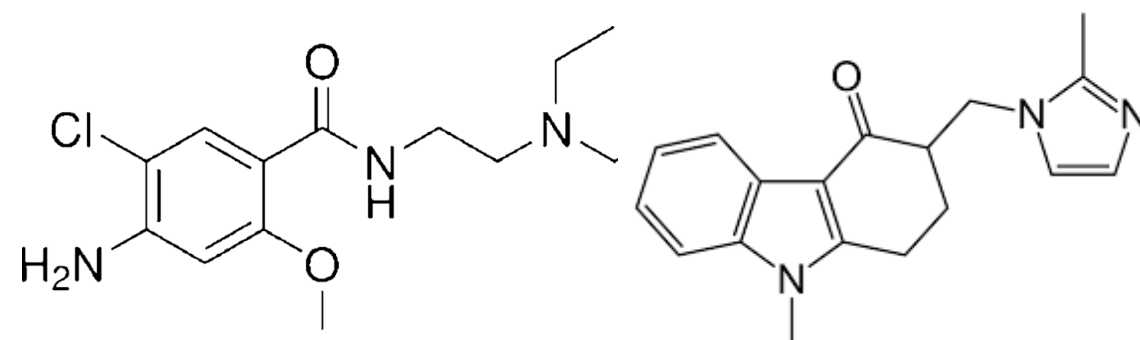
**-เดชา ศิริภัทร
ประธานมูลนิธิข้าวขวัญ**



Cannabis for cancer evaluation



Cannabinoids Project



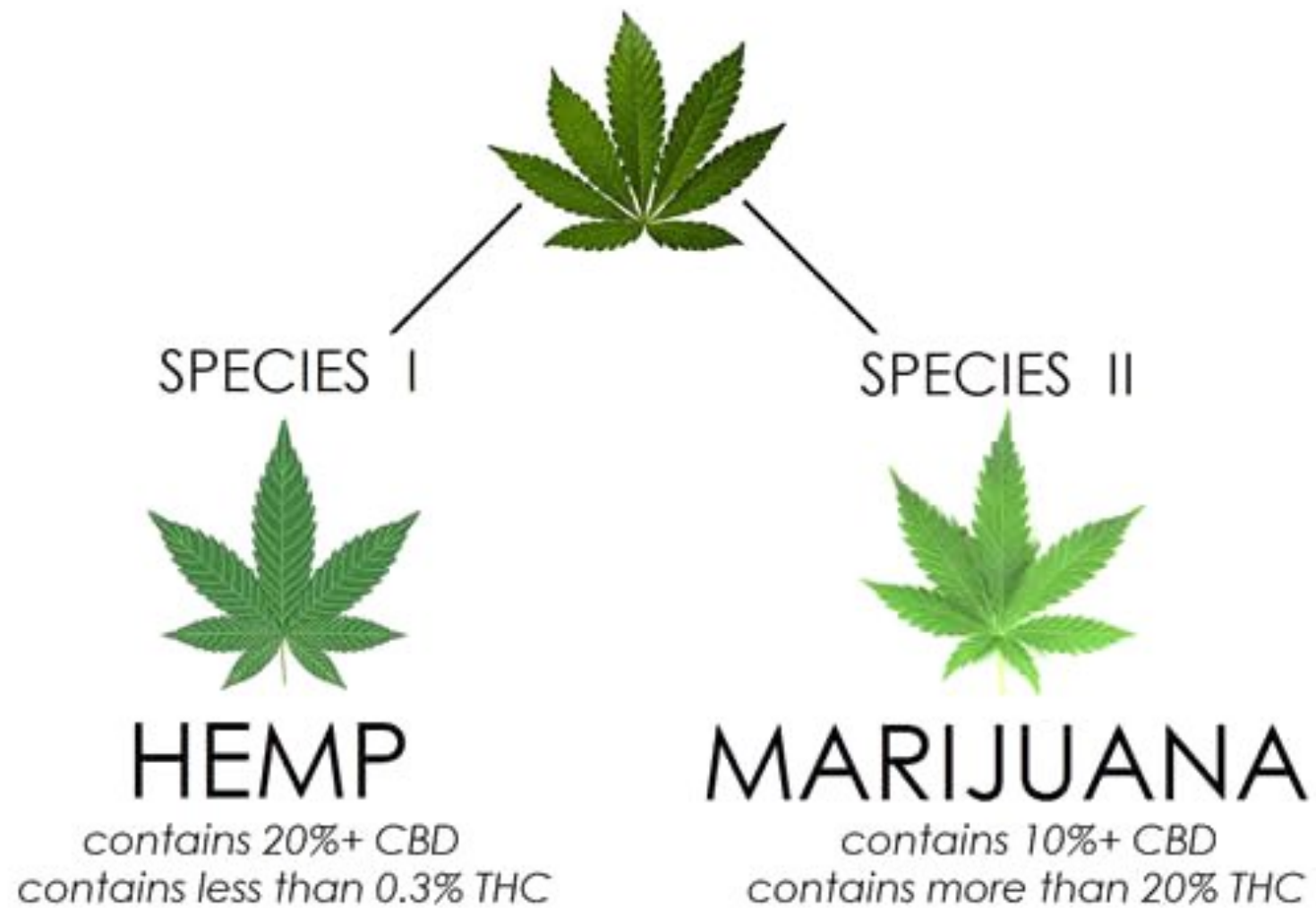
สารสกัดกัญชา ใช้ในสภาวะใดบ้าง
สำหรับผู้ป่วยมะเร็ง ??

ข้อควรระวัง.. ??

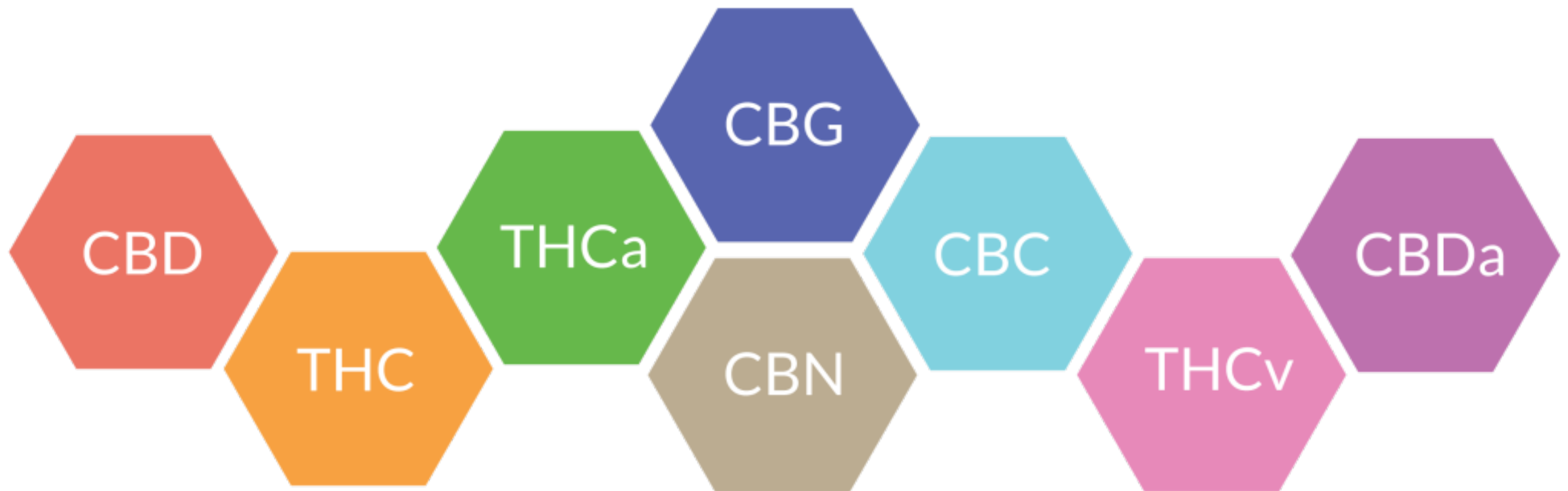
ทำไมต้องมึ่งงานวิจัย เรื่องนี้.??



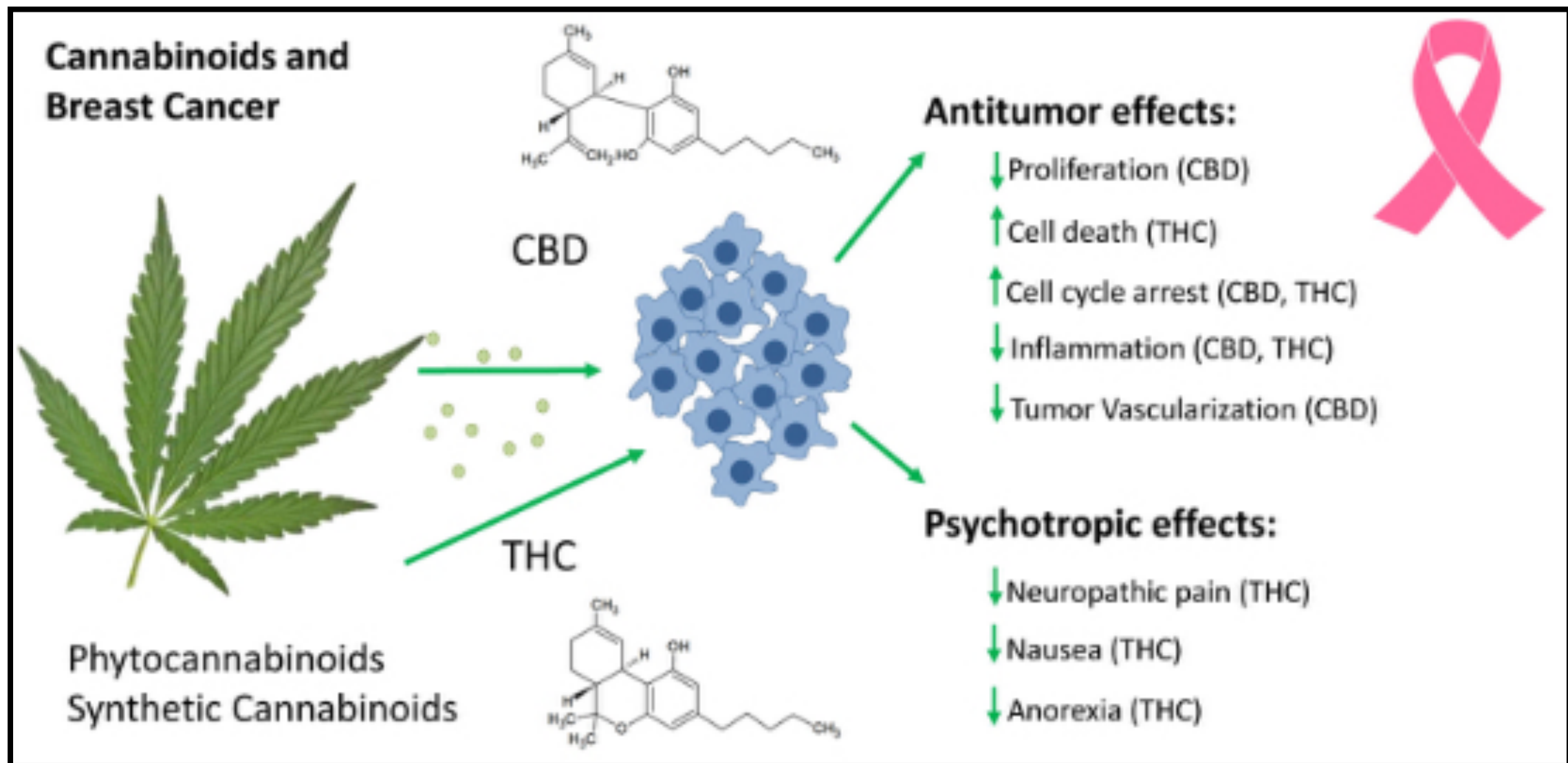
CANNABIS



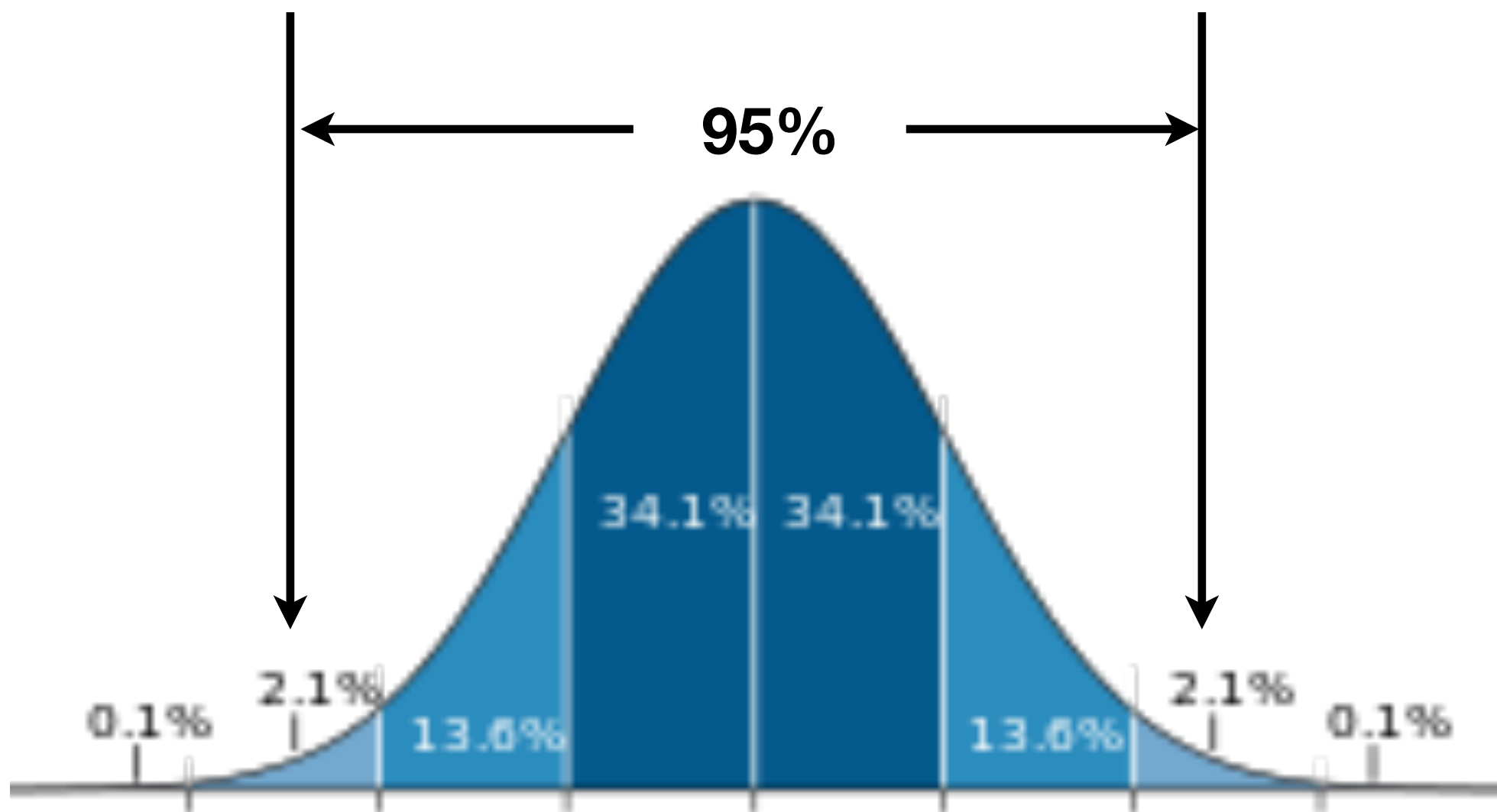
Cannabinoids

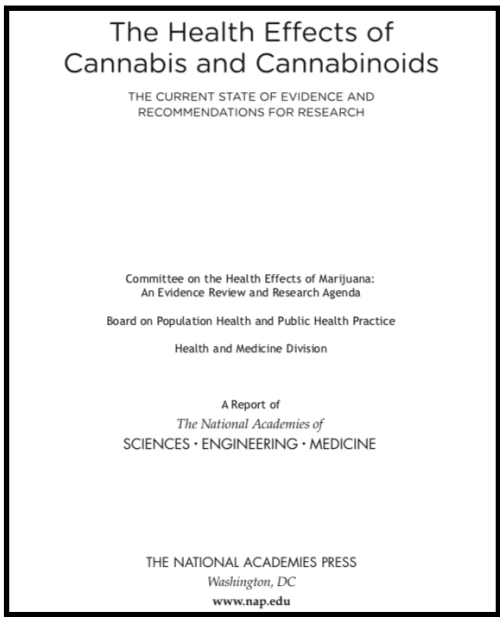


Potential use of Cannabis in Cancer



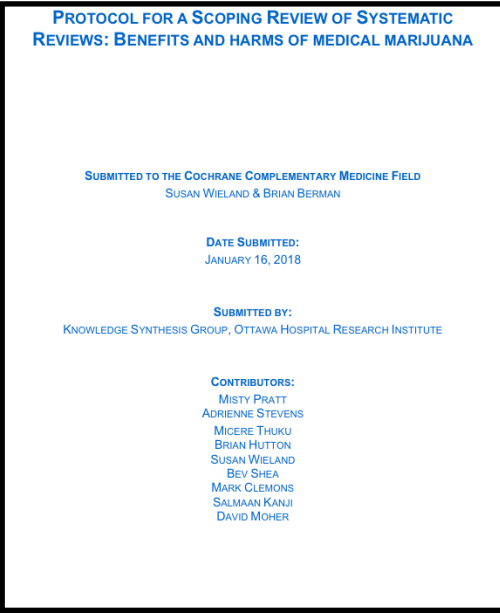
ยาหรือการรักษามาตรฐานที่จะนำมาใช้ทางการแพทย์



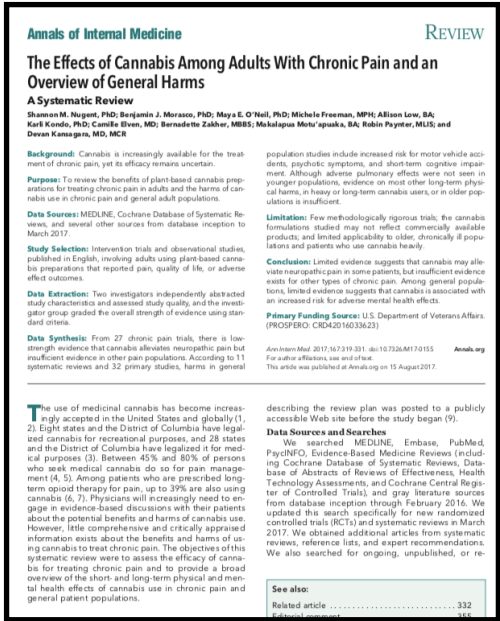


แหล่งข้อมูลที่น่ามาศึกษา

รายงานล่าสุดถึงปี 2019
(1999-2019)



การทบทวนอย่างเป็นระบบ การวิเคราะห์เชิงเปรียบเทียบ รายงานวิจัยแบบสุ่ม รายงานวิจัยแบบการสังเกต



ศักยภาพของกัญชาทางการแพทย์(1)

**‘มีหลักฐาน และข้อมูลเพียงพอ ถึง ประสิทธิภาพ
ในการลดอาการคลื่นไส้อาเจียน จากยาเคมีบำบัด
(สารสกัดกัญชา ชนิดรับประทาน)’**

เชื่อว่า ผ่านกลไก 3(5-HT₃), CB1& GABA receptor

สารสกัด THC & CBD จากต้นกัญชา **สเปรย์พ่นปาก**
(100 microlitre spray: THC 2.7 mg & CBD 2.5 mg)

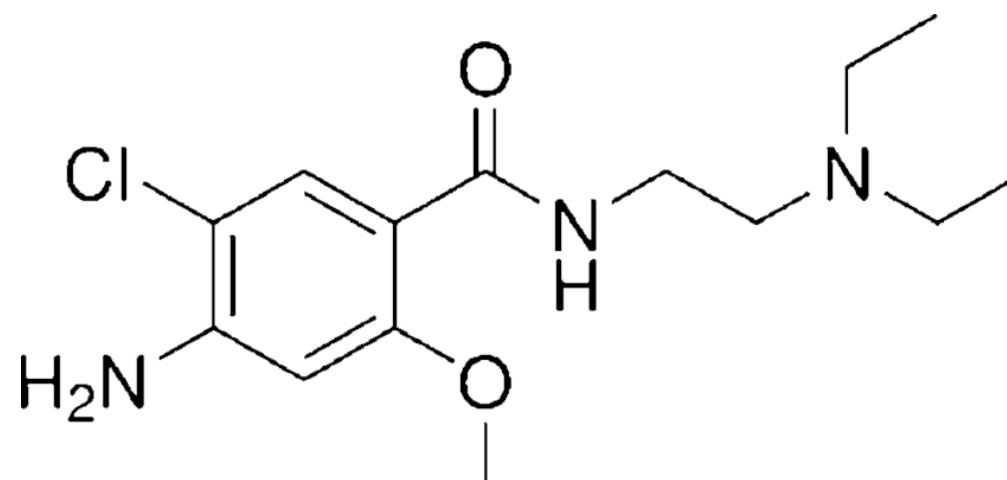
สารสกัดกัญชา & ผลข้างเคียง ระยะสั้น

- เหตุการณ์ไม่พึงประสงค์โดยรวม 3.03 เท่า (OR 3.03 [95%CI 2.42, 3.80])
- รุนแรง (serious) 1.41 เท่า (OR 1.41 [95%CI 1.04, 1.92])
- ผู้ป่วยถอนตัวเนื่องจาก AE 2.94 เท่า (OR 2.94 [95%CI 2.18, 3.96])
- อาการผิดปกติทางระบบประสาท 3.17 เท่า (OR 3.17 [95%CI 2.20, 4.58])
- อาการผิดปกติทางจิต 3.10 เท่า (OR 3.10 [95%CI 1.81, 5.29])
- Disorientation 5.41 เท่า (OR 5.41 [95%CI 2.61, 11.19])
- Dizziness 5.09 เท่า (OR 5.09 [95%CI 4.10, 6.32])
- Euphoria 4.08 เท่า (OR 4.08 [95%CI 4.08, 7.64])
- Confusion 4.03 เท่า (OR 4.03 [95%CI 2.05, 7.97])

ยาลดอาการคลื่นไส้อาเจียน



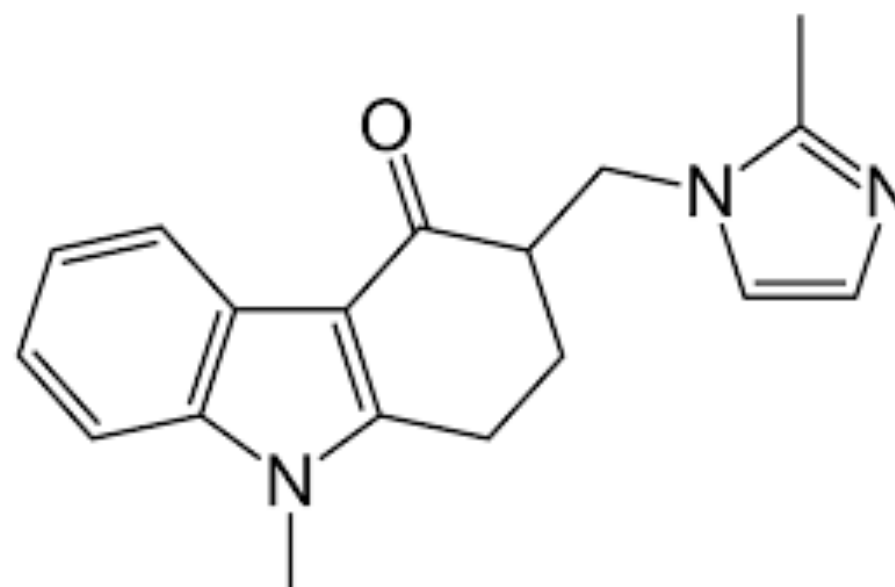
Metoclopramide



C₁₄H₂₂ClN₃O₂



**Ondansetron
(1990/2006)**

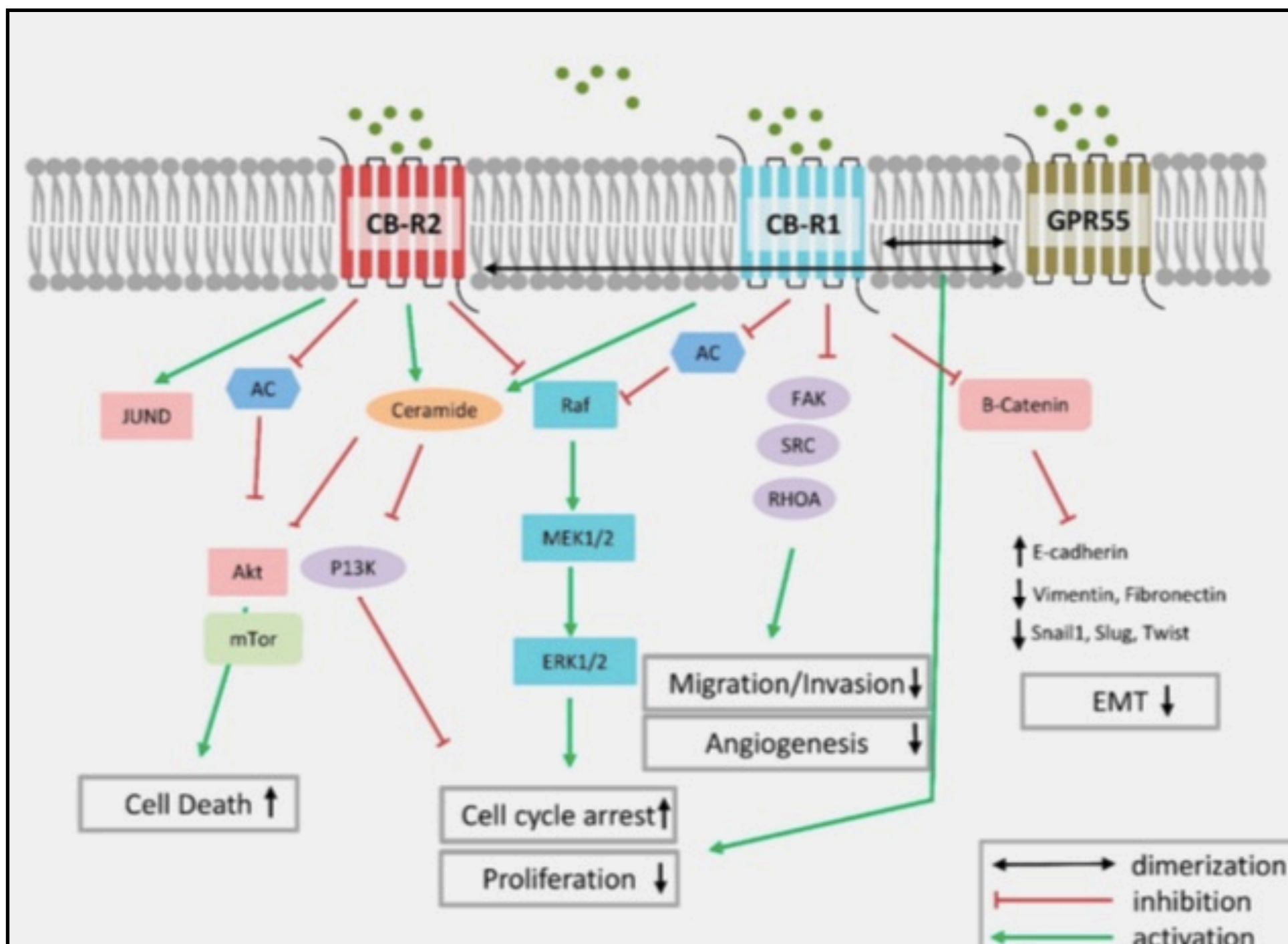


C₁₈H₁₉N₃O

สารสกัดกัญชา กับการรักษามะเร็ง

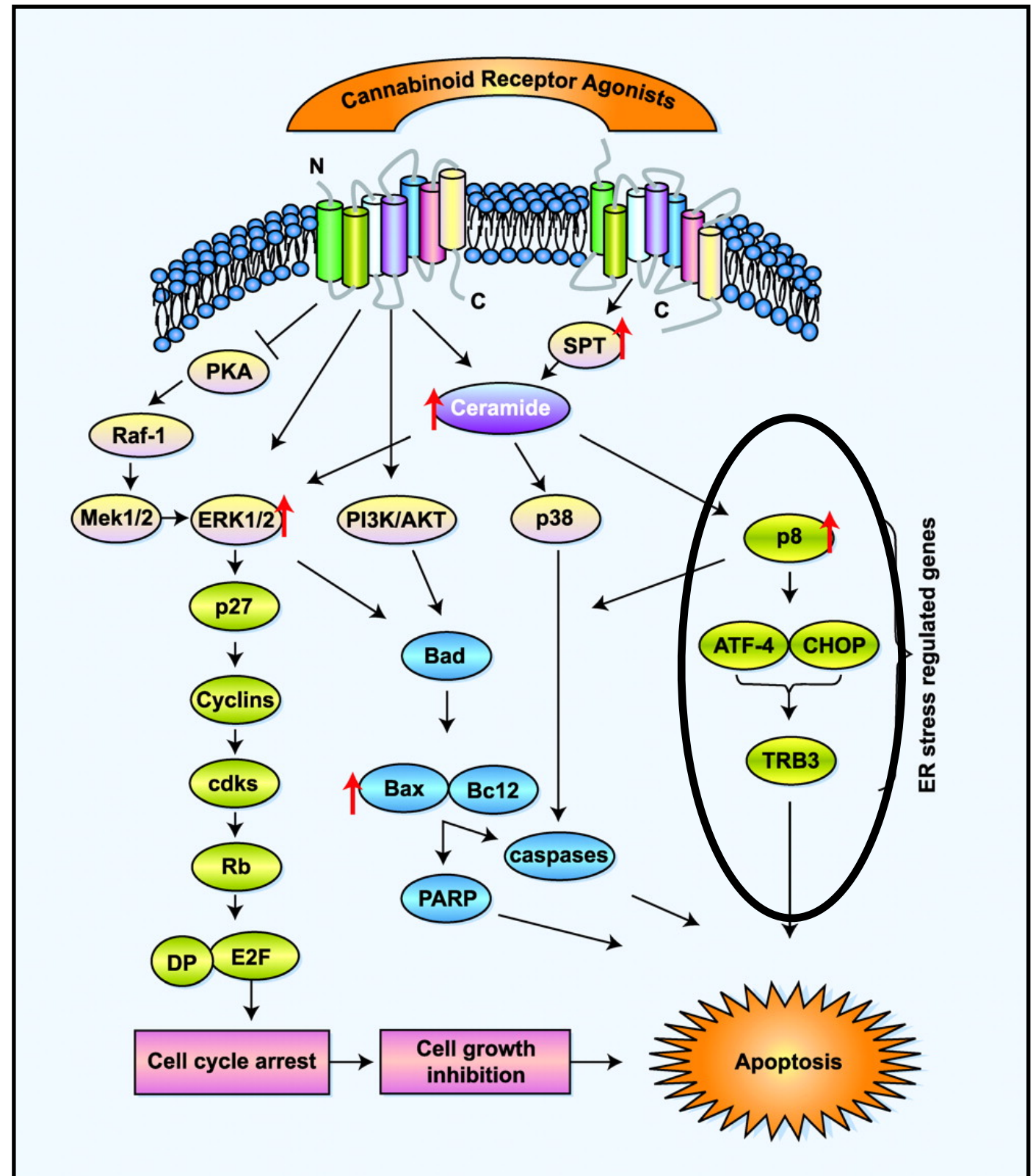


Mechanism of action



Mechanism of action

CB1-R/ CB2-R/ others



Cannabinoids related drugs

<i>Compound</i>	<i>Target(s)</i>	<i>Potential therapeutic applications</i>
CP-55,940	Nonselective agonist (CB1 = CB2)	Analgesic, antiemetic, appetite stimulant, tumour growth inhibitor, multiple sclerosis
WIN 55,212-2	Nonselective agonist (CB1 = CB2)	Analgesic, antiemetic, appetite stimulant, tumour growth inhibitor, multiple sclerosis
HU-210	Nonselective agonist (CB1 = CB2)	Analgesic, multiple sclerosis, neuroprotective
Δ^9 -THC	Nonselective agonist (CB1 > CB2)	Analgesic, antiemetic, appetite stimulant tumour growth inhibitor
Anandamide	Nonselective agonist (CB1 \gg CB2)	Analgesic, antiemetic, appetite stimulant, tumour growth inhibitor
(R)-methanandamide	Nonselective agonist (CB1 \gg CB2) metabolically stable	Analgesic, antiemetic, appetite stimulant, tumour growth inhibitor
2-AG	Nonselective agonist (CB1 > CB2)	Analgesic, antiemetic, appetite stimulant, tumour growth inhibitor
O-1269	Partial CB1 agonist	
Noladin ether	Selective CB1 agonist	Analgesic
AM-1241	Selective CB2 agonists	Tumour growth inhibitor (in glioma, skin carcinoma, lymphoma and leukaemia); multiple sclerosis immune diseases peripheral analgesia
HU-308		
JWH-133		
JWH-015		
BML190	Nonselective agonist (CB2 \gg CB1)	

Effects of Δ^9 -THC and CBD at sub-micromolar concentrations upon cell viability, motility, and invasiveness in human immortalised cell lines

Cell lines and THC/CBD incubation conditions	Compound tested	Findings	References
HeLa cervical cancer cells , serum-free, 72 h	Δ^9-THC	0.01, 0.1, and 1 μ M Δ^9 -THC \downarrow cell invasion index (Boyden chamber ^b). Effect of 1 μ M Δ^9 -THC blocked by AM251 + AM630, by inhibitors of p38 and p42/44 MAP kinase activation, and by siRNA to tissue inhibitor of matrix metalloproteinases-1.	Ramer R & Hinz B (2008) J Natl Cancer Inst. 2008;100:59-69.
LNCaP prostate cancer cells , serum-free, 48 h incubation	Δ^9-THC	\uparrow thymidine incorporation seen with max effect at 0.125-0.25 μ M Δ^9 -THC, thereafter tailing off at 0.5 and 1 μ M. Similar effect seen with R(+)-methAEA and HU-210	Sánchez MG et al. (2003) FEBS Lett.2003;555:561-6.
PC3 prostate cancer cells , serum-free, 48-72 h	Δ^9-THC	50 and 100 nM, but not 250 nM Δ^9 -THC \uparrow thymidine incorporation and cell viability (MTT assay, 72 h). 100 nM Δ^9 -THC \uparrow Akt phosphorylation, Erk phosphorylation, Raf-1 translocation, and EGFR production (48 h). Experiments with rimonabant and SR144528 and suggest involvement of both CB ₁ and CB ₂ receptors. Similar results seen with 100 nM R(+)-methAEA.	Sánchez MG et al. (2003) Cell Signal.2003;15:851-9.
Panc1, Capan2, BxPc3, Mia-PaCa2 pancreatic cancer cell lines; 0-0.1% fetal bovine serum, 66 h	Δ^9-THC	Capan2 \uparrow cell viability (Cell titer 96 assay) at 0.5 & 1 μ M Δ^9 -THC; Panc at 1 μ M alone. No significant effect of 0.5 or 1 μ M Δ^9 -THC on MiaPaCa2 and BxPc3 cell viability. All four cell lines show complete loss of cell viability after incubation with 3 and 4 μ M Δ^9 -THC .	Carracedo A et al. (2006) Cancer Res.2006;66:6748-55.
AV40-transformed human trabecular meshwork cell line, serum-free, 24 h or 72 h	Δ^9-THC	At concentration range 0.1 nM to 1 μ M, no effects of Δ^9 -THC on cell viability (WST-1 assay). Significant \downarrow cell migration (Boyden chamber, wound-healing assay) with 0.01, 0.1, and 1 μ M Δ^9 -THC. Effect Δ^9 -THC on migration accompanied by \uparrow COX-2 activity.	Ramer R & Hinz B (2010) Biochem Pharmacol. 2010;80:846-57.
Granta 519 mantle cell lymphoma line, 24 & 48 h, 0 or 10% serum	Δ^9-THC	Presence f serum, 24 & 48 h: no effect on cell viability (propidium iodide staining) at either time point for THC in range 1.6-310 nM, \downarrow at 6.3 μ M. Absence of serum, 24 h, no effect of 31 nM Δ^9 -THC upon cell proliferation. The cells expressed CB1 receptors.	Richardson SJ et al. (2007) Leuk Lymphoma. 2007;48:1855-7.
U373-MG glioblastoma cell line; NCI-H292 lung carcinoma cell line, serum-free, 6-24 h	Δ^9-THC	0.1 and 0.3 μ M Δ^9 -THC \uparrow thymidine incorporation (U373-MG cells, 18 h) and cell viability (MTT assay, NCI-H292, 24 h). Effect blocked by EGFR inhibitor AG1478. No apoptosis seen at 0.1-1 μ M THC, but a significant increase at 4-10 μ M THC (NCI-H292, 6 h).	Hart S et al. (2004) Cancer Res.2004;64:1943-50.

Effects of Δ^9 -THC and CBD at sub-micromolar concentrations upon cell viability, motility, and invasiveness in human immortalised cell lines

Cell lines and THC/CBD incubation conditions	Compound tested	Findings	References
SF126 glioblastoma cell line; 0.1% fetal bovine serum, 72 h	Δ^9 -THC	\uparrow Cell viability (MTT assay) at 0.1 μ M Δ^9 -THC, thereafter inhibition (IC_{50} value 0.6 μ M).	McAllister SD et al. (2005) J Neurooncol. 2005;74:31-40.
Glioma cells from biopsies from three patients with recurrent glioblastoma multiforme; 0.5% fetal bovine serum, 48 h	Δ^9 -THC	No obvious effect on cell viability (trypan blue exclusion) at 0.5 μ M, but large reduction at 2.5 μ M. Biopsies expressed both CB1 and CB2 receptors, and the effect of 2.5 μ M Δ^9 -THC was blocked by a combination of rimonabant + SR144528.	Guzmám M et al. (2006) Br J Cancer. 2006;95:197-203.
THC-” sensitive” (Gos3, U87, A172, SW1783, U118) and “resistant” (U373, T98, SW1088, CCF, LN405) glioma cell lines, serum-free, 72 h	Δ^9 -THC	Possible \uparrow cell viability (MTT assay) for some cell lines (e.g., T98), not for others (e.g., SW1088) at 0.5 μ M Δ^9 -THC. “Sensitive” lines were completely inhibited at 3 μ M (IC_{50} <2.5 μ M), all “resistant” lines except LN405 (40% reduction) at 4 μ M (IC_{50} >2.5 μ M).	Lorente M et al. (2011) Cell Death Differ. 2011;18:959-73.
CaCo-2 and HCT116 colon adenocarcinoma cells , serum-free, 24 h	CBD	Cell viability: no effect 0.01-10 μ M CBD (neutral red uptake assay); no effect at 0.01-1 μ M, modest reduction at 10 μ M CBD (MTT assay). Thymidine incorporation: Caco-2 cells show \downarrow at 0.1, 1, and 10 μ M, HCT116 cells only at 10 μ M CBD	Aviello G et al. (2012) J Mol Med (Berl). 2012;90:925-34.
Caco-2 colorectal adenocarcinoma cells , 1% or 5% serum, 72 h	CBD	Serum-dependent effects in proliferating cells. At 1% serum, an increased cell viability (XTT assay) was seen at <1 μ M CBD, \downarrow at higher concentration. At 5% serum, only the inhibition was seen. The curves for assays undertaken using physiological O ₂ concentrations were leftward shifted compared with those at atmospheric O ₂ concentrations. No effect of CBD were noted on post-confluent cells. In presence of 0.5 mM butyrate and 8% serum, 0.16 μ M CBD \uparrow cell proliferation (CyQuant assay), Whereas it was blocked by 10 μ M CBD.	Macpherson T et al. (2014) In Vitro Cell Dev Biol Anim. 2014;50:427-26.
Human primary lung carcinoma cells from a brain metastasis, serum-free, 48 h	CBD	Concentration-dependent \downarrow cell viability (WST-1 assay); significant effects seen at all concentrations tested (1 nM – 3 μ M; IC_{50} 0.12 μ M). Maximum effect (~70% reduction in viability) seen at 1 μ M. Effect of 3 μ M reduced by COX-2 and PPAR γ inhibitors. CBD less potent in A549 and H460 lung carcinoma cell (IC_{50} 3.5 and 2.8 μ M, respectively).	Ramer R et al. (2013) Mol Cancer Ther. 2013;12:69-82.
DLD-1 and HCT 116 epithelial colon adenocarcinoma cells , serum-free, 24 h	CBD	No significant effect on cell viability (MTT assay) at 0.3, 1 μ M, \downarrow at 5 μ M (and 3 μ M for DLD-1 cells).	Romano B et al. (2014) Phytomedicine. 2014;21:631-9.

Cell lines and THC/CBD incubation conditions	Compound tested	Findings	References
U87MG glioma cell line, serum-free, 24 h	CBD	No effect of 0.5 μ M CBD, large \downarrow at 1-12 μ M upon cell invasion (BioCoat Matrigel invasion chamber).	Solinas M et al. (2013) PLoS One.2013;8:e76918.
RPMI 8226 and U266 myeloma cells, 10% serum, 72 h	CBD	No effect upon cell viability (MTT assay) at 0.3 and 1 μ M, \downarrow at higher concentrations.	Morelli MB et al. (2014) Int J Cancer.2014;134:2534-46.
Kaposi sarcoma-associated herpes virus-infected human dermal microvascular endothelial cells, 0.5% BSA, 24 h	CBD	Concentration-dependent apoptosis (in situ cell death detection kit), seen at CBD concentrations \geq 0.25 μ M. Apoptosis in infected cells greater than in uninfected cells at CBD \leq 1 μ M. Same level of apoptosis (~75%) for infected and uninfected cells at 2-10 μ M CBD. Cell proliferation (Cell titer 96 assay): no effect of 1 μ M, \downarrow at higher concentrations, with greater effect on infected cells than uninfected cells.	Maor Y et al. (2012) Genes Cancer.2012;3:512-20.
MDA-MB231 breast cancer cell line; 0.1% fetal bovine serum, 72 h (cell viability); serum-free, 20 h (invasivity)	Δ^9 -THC, CBD	\downarrow cell viability (MTT, 72 h): IC ₅₀ values TCA 1.2 μ M, CBD 1.3 μ M. Invasivity (Boyden chamber, 20 h): no effect of Δ^9 -THC at 0.1 or 1 μ M, ~50% reduction at 1.5 μ M; ~40, ~60, and ~90% reduction at 0.1, 1, and 1.5 μ M CBD. CBD effect parallels reduction of 1d-1 expression, and ectopic expression of this protein blocks CBD invasivity.	McAllister SD et al. (2007) Mol Cancer Ther.2007;6:2921-7.
SF126, U251, and U87 glioblastoma, 72 h	Δ^9 -THC, CBD	Cell viability (MTT assay): IC ₅₀ values 2.5-3.3 μ M (Δ^9 -THC) and 0.6-1.2 μ M (CBD). In U251 cells, two-way ANOVAs indicated significant interactions CBD + Δ^9 -THC for combination of IC ₂₀ and IC ₈₀ concentration. 1.7 μ M Δ^9 -THC + 0.4 μ M CBD: \uparrow annexin-positive cells, reduced by SR144528, blocked by α -tocopherol. Combination also \uparrow reactive oxygen species. Cell invasion (Boyden chamber): U251 cell IC ₅₀ values ~85 nM (Δ^9 -THC), ~130 nM (CBD). No synergistic effects reported for combination of 100 nM.	Marcu JP et al. (2010) Mol Cancer Ther.2010;9:180-9.
U87MG glioma, serum-free, 72 h	Δ^9 -THC, CBD	IC ₅₀ values for inhibition of cell viability (MTT assay) 1.2 μ M (Δ^9 -THC) and 1.5 μ M (CBD). No significant increase in cell viability seen at low concentrations. Combination of 0.9 μ M THC + 0.9 μ M CBD \downarrow cell viability ~20%, whereas compounds per se had no significant effect. Similar result for SW1783 cells and for combination of Δ^9 -THC with tomozolomide (25-75 μ M). Effect of Δ^9 -THC / CBD combination not seen in cells where autophagy or apoptosis is compromised.	Torres S et al. (2011) Mol Cancer Ther.2011;10:90-103.
Human T98G, U87MG, and mouse GL261 glioma cells, 5% serum, 48 h	THC, CBD and botanical preparations	No obvious effect (mean values in the range 90-115% of control) of the compounds at 0.5 or 1 μ M upon cell proliferation (resazurin reduction assay). The compounds were inhibitory at higher concentrations	Scott KA et al. (2014) Mol Cancer Ther. 2014;13:2955-67.

CB	Cell Line	IC ₅₀	Antitumoral Activity	Receptor Mechanism	Citation
THC	MDA-MB-231	5.0 ± 1.2 µM	Induction of apoptosis Cell cycle arrest, Inhibition of G2-M transition via downregulation of Cdc2	CB2-R	[44]
	MDA-MB-468	4.4 ± 0.3 µM			
	SKBR-3	4.5 ± 0.4 µM			
	MCF-7	10.2 ± 0.7 µM			
	EVSA-T	4.0 ± 0.1 µM			
THC	T-47D	6.7 ± 0.2 µM	Increased production of IL-4 and IL-10 Suppression of the cell-mediated Th1 response and enhancement of the Th2-response	CB1-R CB2-R	[45]
	MDA-MB-231	n.d.			
	MCF-7	n.d.			
	4T1	n.d.			
THCA	MDA-MB-231	1.2 µmol/L	Antiproliferative activity Reduction of invasiveness via ID-1	n.d.	[46]
	MDA-MB-468	2.5 µmol/L			
	MCF-7	14.2 ± 2.1 µM			
	MDA-MB-231	24.3 ± 4.2 µM			
	MCF-7	9.8 ± 0.4 µM			
CBD	MDA-MB-231	18.2 ± 5.3 µM	Inhibition of cell viability Induction of apoptosis/ autophagy No influence on cell viability	CB2-R	[47]
	MCF-7	n.d.			
	SK-BR-3	n.d.			
	ZR-75-1	n.d.			
	MCF-10A (n.m.)	n.d.			
CBD	MDA-MB-231	8.2 ± 0.3 µM	Inhibition of cell viability Cell cycle arrest at the G1/S transition Induction of apoptosis via pro-caspase-3 cleavage to caspase-3, induction of endoplasmic reticulum stress, inhibition of mTOR and Akt	CB2-R	[47]
	MCF-7	10.6 ± 1.8 µM			
	MDA-MB-231	2.2 µM			
	T-47D	5.0 µM			
	MDA-MB-231	1.3 µmol/L			
CBDA	MDA-MB-436	1.6 µmol/L	Antiproliferative activity Invasiveness reduction via ID-1	n.d.	[46]
	MDA-MB-231	>100 µM			
	MDA-MB-231	25 µM			
	MDA-MB-231	>25 µM			
	MCF-7	21.7 ± 3.2 µM			
CBN	MDA-MB-231	>25 µM	Inhibition of cell growth	CB2-R	[47]
	MDA-MB-231	>25 µM			
CBN	MDA-MB-231	1.2 µmol/L	Antiproliferative activity Invasiveness reduction via ID-1	n.d.	[46]
	MDA-MB-468	2.6 µmol/L			

CB	Cell Line	IC ₅₀	Antitumoral Activity	Receptor Mechanism	Citation
CBG	MDA-MB-231	2.3 µmol/L	Antiproliferative activity	n.d.	[46]
	MDA-MB-468	2.1 µmol/L	Invasiveness reduction via ID-1		
	MCF-7	9.8 ± 3.4 µM	Inhibition of cell growth	CB2-R	[47]
	MDA-MB-231	20.4 ± 2.6 µM			
CBC	MCF-7	142 ± 1.4 µM	Inhibition of cell growth	CB2-R	[47]
	MDA-MB-231	>25 µM			
AEA	MDA-MB-231	n.d.	No growth inhibition <10 µM	CBI-R	[53]
	MCF-7	0.5 µM	Cell cycle arrest, inhibition of G1/S transition	CBI-R	[54]
	EPM-19	15 ± 0.3 µM			
	BT-474	1.9 µM			
	T-47D	1.9 µM			
	MCF-7	1.4 ± 0.9 µM	Inhibition of adenylyl cyclase and activation of MAPK, thereby exerting a downregulation of PRLr and trk	n.d.	[55]
	EPM-19	15 ± 0.3 µM			
	MCF-7	1.4 ± 0.9 µM	Inhibition of proliferation, inhibition of forskolin-induced cAMP formation, stimulation of RAF1 translocation and MAPK activity	CBI-R CB2-R TRPV	[56]
	T-47D	1.9 ± 0.2 µM			
2-AG	MDA-MB-231	n.d.	Regulation of lipid rafts	CBI-R	[57]
	EPM-19	n.d.	Cell cycle arrest, inhibition of G1/S transition	CBI-R	[54]
	MCF-7	1.4 ± 0.3 µM	Inhibition of proliferation, inhibition of forskolin-induced cAMP formation, stimulation of RAF1 translocation and MAPK activity	CBI-R CB2-R TRPV	[56]
	T-47D	5.0 ± 1.1 µM			
	MDA-MB-231	n.d.	Inhibition of adhesion and migration on type IV collagen without modifying integrin expression	CBI-R	[53]
	T-47D	n.d.			
Met-F-AEA	MDA-MB-231	n.d.	Inhibition of proliferation by degradation of b-catenin and decrease in cyclin D1, c-Myc and MMP-2	CBI-R	[58]
	MDA-MB-231	n.d.	Cell cycle arrest, inhibition of G1/S transition		
	MDA-MB-231	n.d.	Upregulation of E-cadherin accompanied by the reduction of vimentin, fibronectin and N-cadherin	n.d.	[59]
	MDA-MB-231	n.d.	Inhibition of angiogenesis by the reduction of pro-angiogenic factors VEGF		
	MDA-MB-231	n.d.	Reduction of metalloproteinases, TIMP1 and TIMP2		
(R)-Met-AEA	EPM-19	0.8 µM	Cell cycle arrest, inhibition of G1/S transition	CBI-R	[54]

Research Paper

CB₂ cannabinoid receptor activation promotes colon cancer progression via AKT/GSK3 β signaling pathway**Esther Martínez-Martínez¹, Asunción Martín-Ruiz¹, Paloma Martín², Virginia Calvo¹, Mariano Provencio¹, José M. García¹**¹Department of Medical Oncology, Hospital Universitario Puerta de Hierro-Majadahonda, E-28222 Madrid, Spain²Department of Pathology, Hospital Universitario Puerta de Hierro-Majadahonda, E-28222 Madrid, Spain

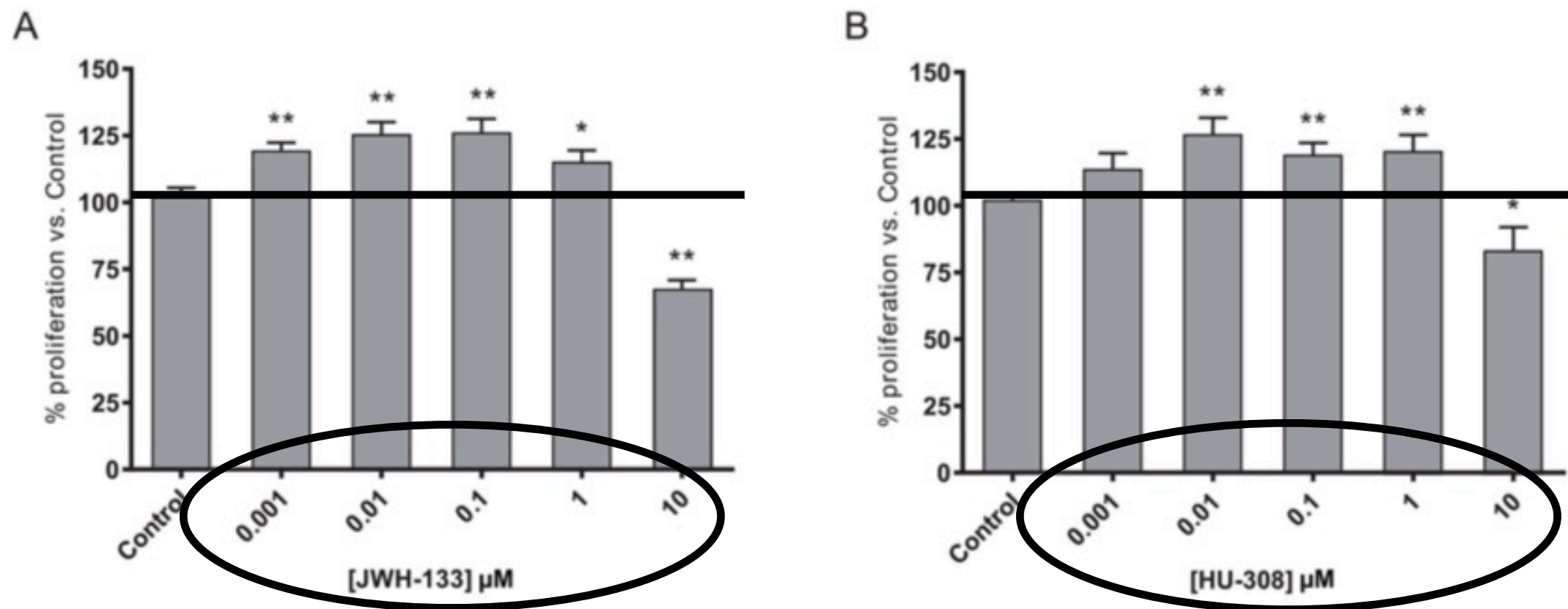
Correspondence to: José M. García, email: jmgarcia@idiphim.org

Keywords: CB₂, colon cancer, AKT/PKB, JWH-133, proliferation

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**“Dose dependence response”**

Appraising the “entourage effect”: Antitumor action of a pure cannabinoid *versus* a botanical drug preparation in preclinical models of breast cancer

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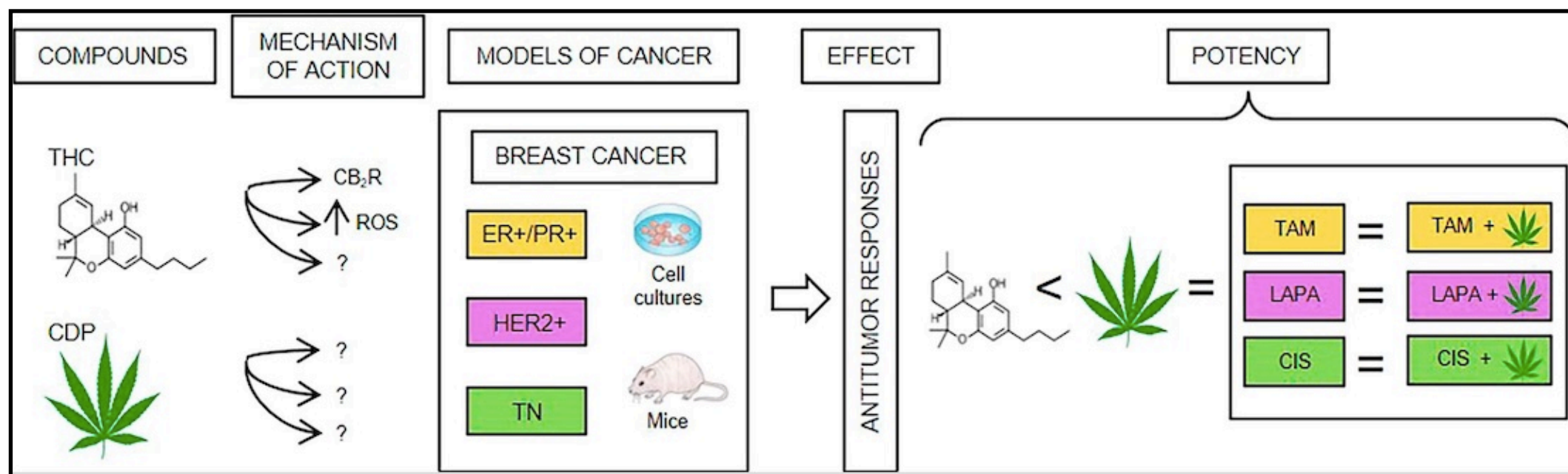
^a Complutense University, Madrid, Spain

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^d Instituto Ramón y Cajal de Investigación Sanitaria, CIBERNED and IUIIN, Madrid, Spain



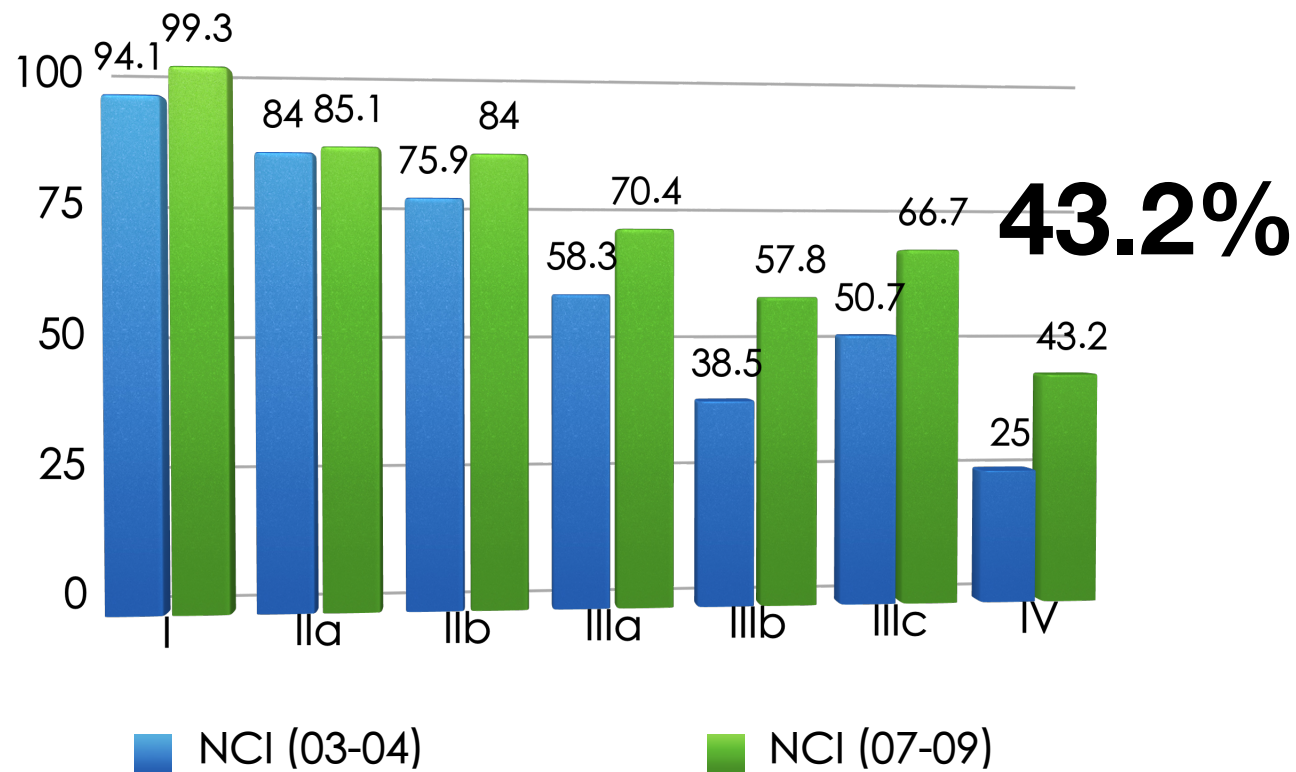


“The combination of cannabinoids with estrogen receptor- or HER2-targeted therapies (tamoxifen and lapatinib, respectively) or with cisplatin, produced **additive anti-proliferative** responses in **cell cultures**”

“Combinations of these treatments **in vivo** showed **no interactions**, either positive or negative”

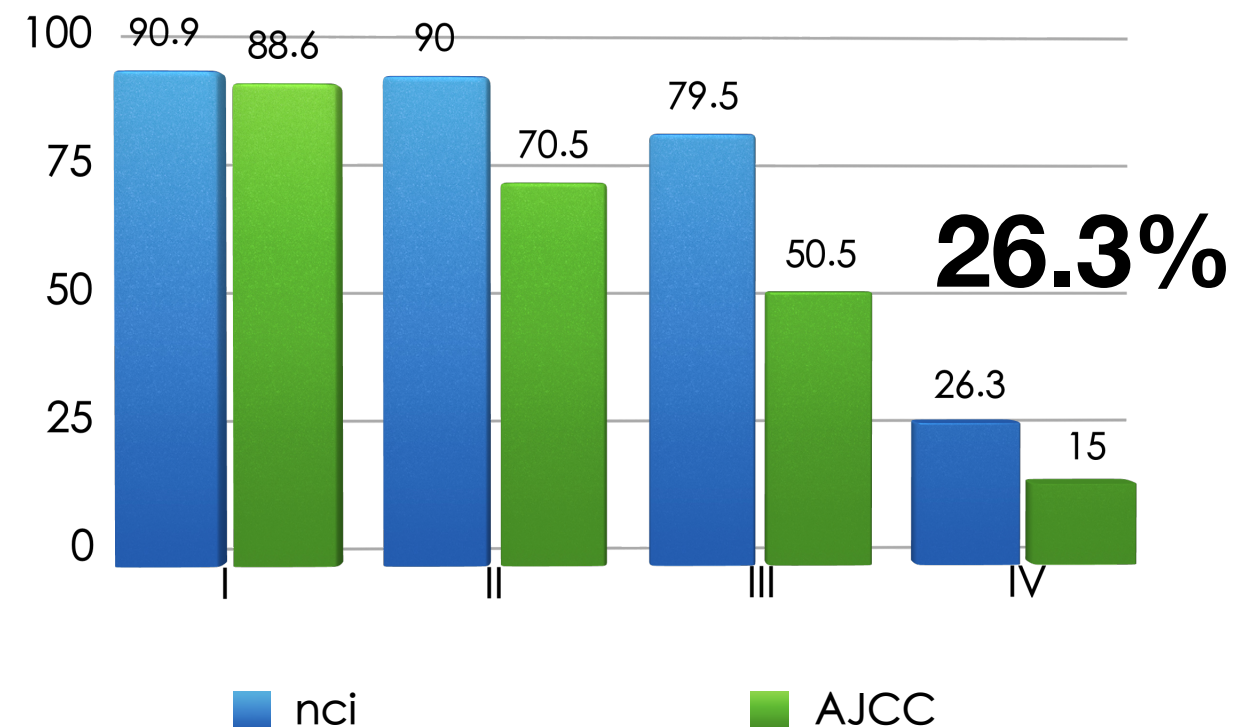
อัตราการอยู่รอดชีพ 5 ปี ในมะเร็งชนิดต่างๆ ด้วยการรักษาปัจจุบัน

99.3%



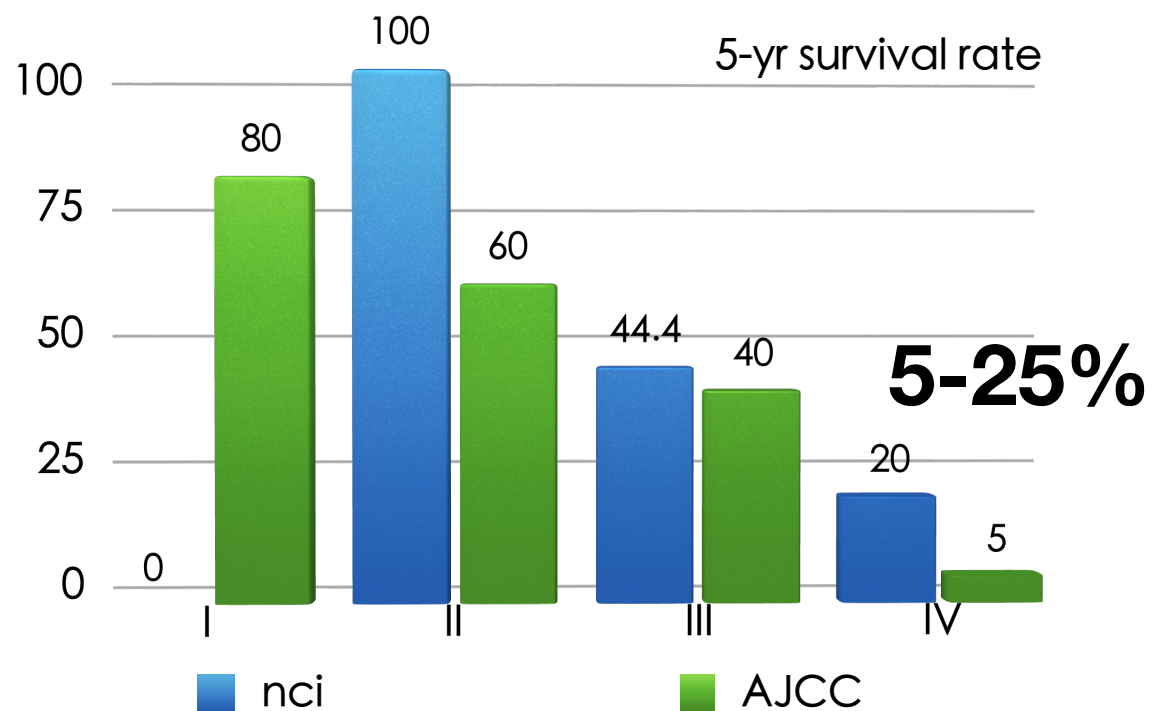
มะเร็งเต้านม

90.9%



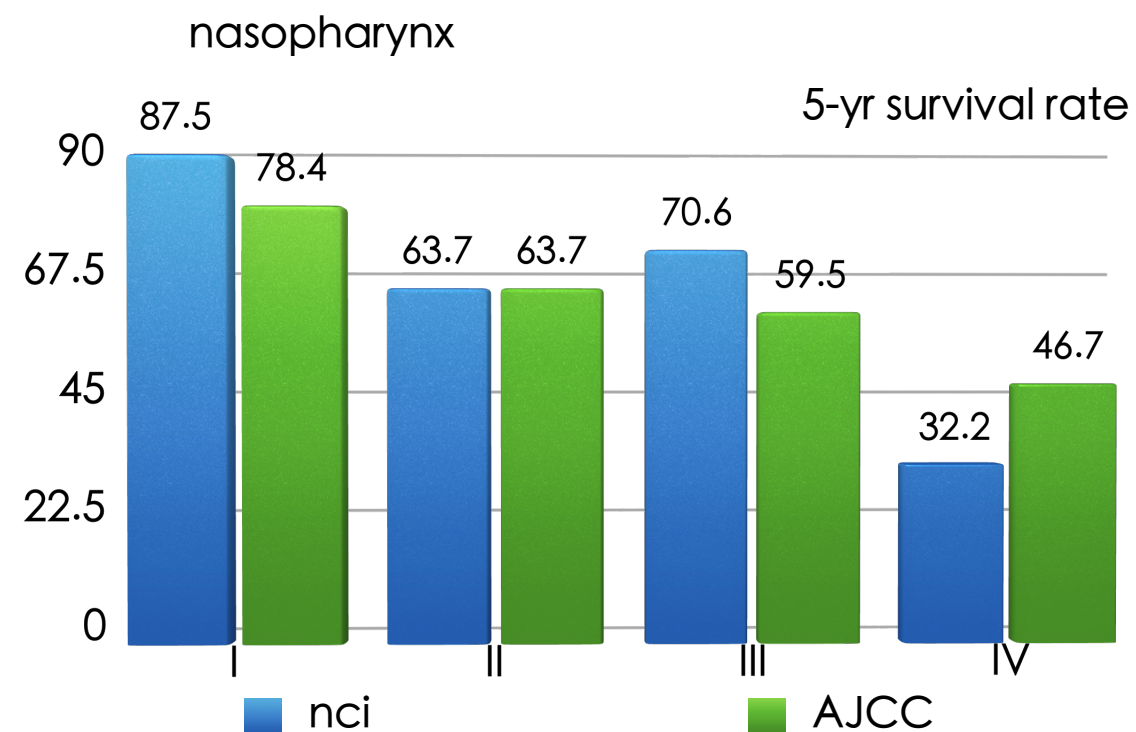
มะเร็งลำไส้ใหญ่

80-100%



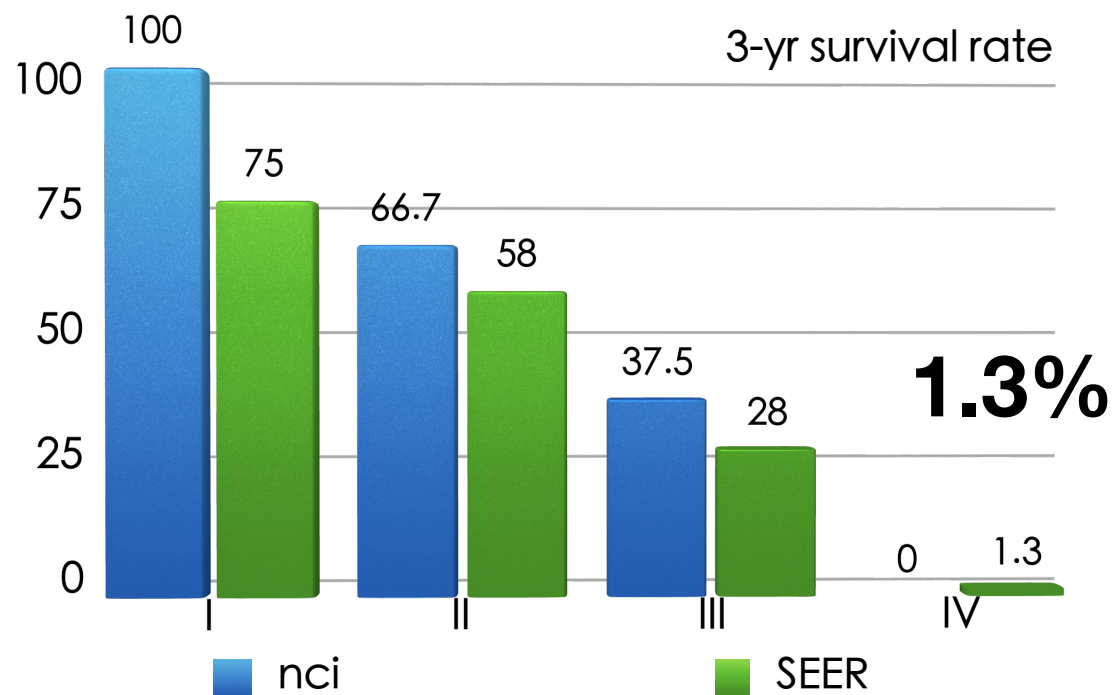
มะเร็งกระเพาะอาหาร

87.5%



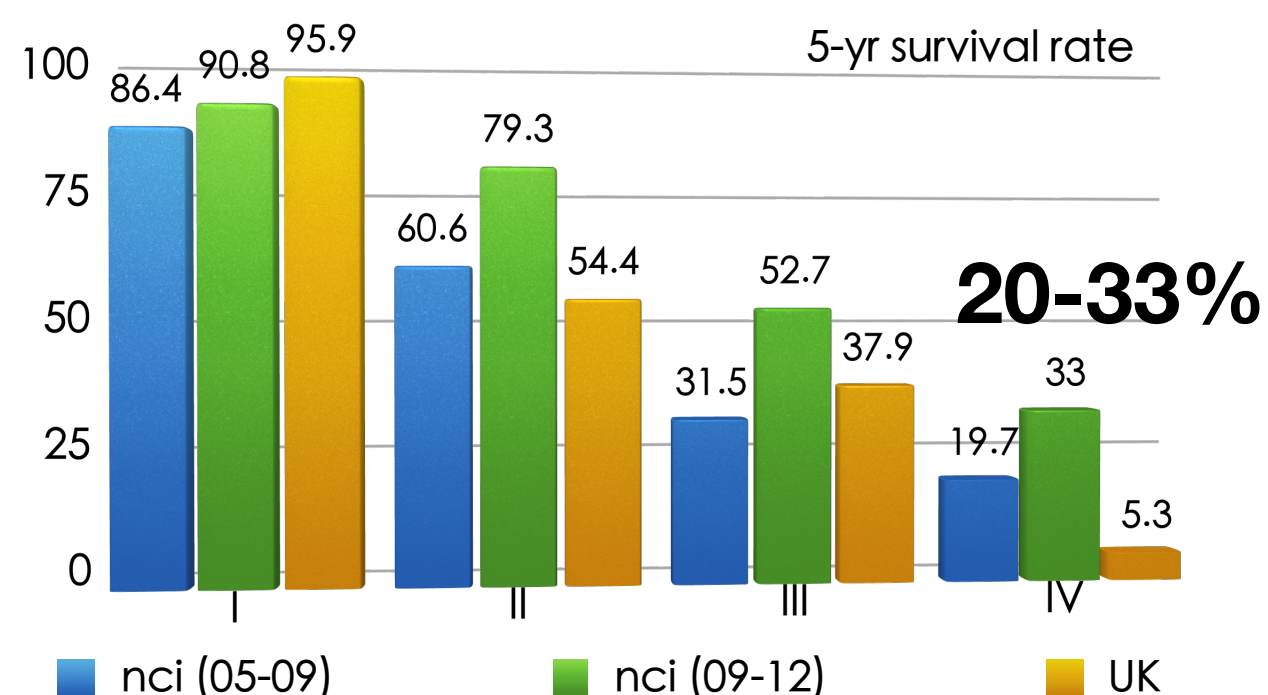
มะเร็งหลังโพรงจมูก

75-100%



มะเร็งตับ

91-96%



มะเร็งปากมดลูก

ศักยภาพของกัญชาทางการแพทย์(2)

**‘ไม่มีหลักฐาน และข้อมูลเพียงพอ ที่จะสนับสนุนถึง
ประสิทธิผลของสารสกัดกัญชาในการรักษามะเร็ง**

ศักยภาพของกัญชาทางการแพทย์(2)

**‘มีหลักฐานจำนวนมาก ถึง ประสิทธิภาพ
ของสารสกัดกัญชาต่อเซลล์มะเร็งหลายชนิด
ในระดับหลอดทดลอง และสัตว์ทดลอง’**

เชื่อว่า ผ่านกลไก ทั้ง CB และ non-CB receptor

มีความหลากหลายของส่วนประกอบ THC & CBD ในแต่ละการวิจัย

ศักยภาพของกัญชาทางการแพทย์(2)

**‘ควรมีการพัฒนางานวิจัยทั้งทางระดับหลอดทดลอง
สัตว์ทดลองและมนุษย์ เพื่อหาสัดส่วนของ
สารสกัดกัญชาที่เหมาะสมต่อการรักษามะเร็ง’**

สารสกัดกัญชา กับ อาการปวดจากมะเร็ง



ศักยภาพของกัญชา ทางทางการแพทย์(3)

‘ไม่มีหลักฐานสนับสนุนเพียงพอ หรือคัดค้าน ถึง
ประสิทธิผลในการใช้ในการรักษา**อาการปวด**
สำหรับผู้ป่วยมะเร็ง

Nabiximols (THC:CBD =1:1) : ขึ้นทะเบียนครั้งแรกเป็นยาแก้ปวดสำหรับ
ผู้ป่วยมะเร็งในสหรัฐอเมริกา

ศักยภาพของกัญชาทางการแพทย์(3)

**‘ไม่มีหลักฐานสนับสนุนเพียงพอ หรือคัดค้าน ถึง
ประสิทธิผลในการใช้ในการรักษา **อาการกินไม่ได้**
และผอม สำหรับผู้ป่วยมะเร็งระยะสุดท้าย**

ควรพัฒนางานวิจัยเพื่อหาข้อสรุป รวมทั้งหาสัดส่วนสารสกัดกัญชาที่เหมาะสม



dr.somchai.t@gmail.com